

**Supplemental Material**

**E2F1-Mediated *FOS* Induction in Arsenic Trioxide–Induced  
Cellular Transformation: Effects of Global H3K9  
Hypoacetylation and Promoter-Specific Hyperacetylation *in Vitro***

Sunniyat Rahman, Zjwan Housein, Aleksandra Dabrowska, Maria Dolores Mayán, Alan R.  
Boobis, and Nabil Hajji

## **Plasmids**

Plasmids used were HDAC1 (Addgene # 13820), HDAC3 (Addgene # 13819) and HDAC4 (Addgene # 13821). HDAC2 was generously provided by Dr. Ito Kazuhiro (Imperial College London). PCAF, HMOF and TIP60 were kindly provided by Dr. Bertrand Joseph (Karolinska Institutet).

## **Antibodies**

For histone acetylation analysis the following antibodies were used: anti-acetyl Histone H3-Lys 9, anti-acetyl Histone H4-Lys 12 and anti-acetyl Histone H4-Lys 16 (all from Millipore). For intracellular protein analysis the following antibodies were used: anti-PARP, Mdm2, p53 phosphorylated serine 15, p53, Bid, Caspase3 and  $\beta$ -Actin (all from Cell Signalling).

**Table S1.** Primers used for qRT-PCR.

<b>Target</b>	<b>Forward 5'-&gt; 3'</b>	<b>Reverse 5'-&gt; 3'</b>
<i>cFOS</i>	TCGGGCTTCAACGCAGACTACG	AAGGAGTCTGCGGGTGAGTGGT
<i>cJUN</i>	AAGCGCATGAGGAACCGCATCG	TCACTTTTTCTCCAGCCGGGC
<i>MDM2</i>	TTCCCAGCCTAGGTTTCAGA	AACACGGAGCTTGAGAGGAA
<i>P53</i>	GTGGTTTCAAGGCCAGATGT	GGCCCACTTCACCGTACTAA

**Table S2.** Primers used for ChIP.

<b>Target Promoter</b>	<b>Forward 5'-&gt; 3'</b>	<b>Reverse 5' -&gt; 3'</b>
<i>P53</i>	CAGTTGCAAACCAGACCTCA	GTGGAAGGAAATTTGCGTGT
<i>BAX</i>	GAGACACTCGCTCAGCTTCTT	TTCATCCAGGATCGAGCAG
<i>PUMA</i>	GGACAGTCGGACACACACAC	GTACATCCTCTGGGCTCTGC
<i>C-MYC</i>	TGGCGGGAAAAAGAACGGAG	GAAGCCGCTCCACATACAGT
<i>C-FOS</i>	CAGACTACGAGGCGTCATCC	AGTTGGTCTGTCTCCGCTTG